## ACS Medicinal Chemistry Letters

Letter

Subscriber access provided by Kaohsiung Medical University

## Novel aminoquinoline derivatives significantly reduce parasite load in Leishmania infantum infected mice

Jelena Konstantinovi#, Milica Videnovic, Stefania Orsini, Katarina Bogojevic, Sarah D'Alessandro, Diletta Scaccbarozzi, Natasa Terzic Jovanovic, Luigi Gradoni, Nicoletta Basilico, and Bogdan A. Solaja ACS Med. Chem. Lett., Just Accepted Manuscript • DOI: 10.1021/acsmedchemlett.8b00053 • Publication Date (Web): 04 May 2018 Downloaded from http://pubs.acs.org on May 6, 2018

### **Just Accepted**

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

7

8 9

10

11 12

13 14

15

16

17

18 19

20

21

22 23

24

25

26

27

28

29

30 31 32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57 58 59

60

# Novel aminoquinoline derivatives significantly reduce parasite load in *Leishmania infantum* infected mice

Jelena Konstantinović<sup>¶</sup>, Milica Videnović<sup>#</sup>, Stefania Orsini<sup>†</sup>, Katarina Bogojević<sup>¶</sup>, Sarah D'Alessandro<sup>¥</sup>, Diletta Scaccabarozzi<sup>Ψ</sup>, Nataša Terzić Jovanović<sup>⊽</sup>, Luigi Gradoni<sup>†</sup>, Nicoletta Basilico<sup>\*,¥</sup>, Bogdan A. Šolaja<sup>\*,¶,§</sup>

<sup>¶</sup>University of Belgrade, Faculty of Chemistry, Studentski trg 12-16, P.O. Box 51, 11158, Belgrade, Serbia

<sup>#</sup>Faculty of Chemistry Innovative Centre, Studentski trg 12-16, 11158 Belgrade, Serbia

<sup>v</sup>University of Belgrade, Institute of Chemistry, Technology, and Metallurgy, Njegoševa 12, 11000 Belgrade, Serbia

<sup>§</sup> Serbian Academy of Sciences and Arts, Knez Mihailova 35, 11158 Belgrade, Serbia

<sup>¥</sup>Dipartimento di Scienze Biomediche, Chirurgiche e Odontoiatriche, Università degli Studi di Milano, Milan, Italy

<sup>Ψ</sup> Dipartimento di Scienze Farmacologiche e Biomolecolari, Università degli Studi di Milano, Milan, Italy

<sup>†</sup>Unit of Vector-borne Diseases, Istituto Superiore di Sanità, Rome, Italy

KEYWORDS: Leishmania infantum, promastigote, amastigote, mice model, aminoquinoline

**ABSTRACT**: In this paper a detailed analysis of thirty 4-aminoquinoline-based compounds with regard to their potential as antileishmanial drugs has been carried out. Ten compounds demonstrated  $IC_{50}<1 \mu M$  against promastigote stages of *L. infantum* and *L. tropica*, and five compounds showed  $IC_{50}<1 \mu M$  against intramacrophage *L. infantum* amastigotes. Two compounds showed dose-dependent enhancement of NO and ROS production by bone marrow-derived macrophages and remarkable reduction of parasite load *in vivo*, with advantage of being short-term and orally active. To the best of our knowledge, this is the first example of 4amino-7-chloroquinoline derivatives active in *Leishmania infantum* infected mice.

Leishmaniasis is a neglected parasitic disease transmitted by more than 90 sand fly species. The disease may occur in humans and animals, including dogs and rodents. Human infection is caused by about 21 of the 30 species of Leishmania parasites that infect mammals.<sup>1,2</sup> Currently, 2 million people are infected every year and more than 350 million people are at risk mostly in tropical and subtropical areas.<sup>3</sup> The life cycle of Leishmania parasite includes sand fly and vertebrate host stages. After an infected sand fly deposits metacyclic promastigotes into the host's skin during blood feeding, they are phagocytized by macrophages and then transformed into aflagellated amastigotes. In macrophages, amastigotes multiply and after being released they infect new macrophages. The sand fly ingests infected macrophages during a blood meal and the life cycle continues within the sand fly gut.<sup>2</sup> Treatment of leishmaniasis varies and adapts to the severity of the disease and species of the parasite.<sup>1</sup> Antileishmanial drugs that are presently in use are pentavalent antimonials, amphotericin B, pentamidine, miltefosine and others, with AmBisome® and sodium stibogluconate – paromomycin combination therapy being the first choice for fighting leishmania infection (Chart 1).<sup>3,4</sup> All current therapies present serious side effects, including toxicity, and for some of them resistance development are emerging.<sup>5</sup> In addition, the vaccine for preventing human leishmaniasis, which could have an immense influence on suppression of the disease, is still not available.<sup>6</sup> Thus, the development of potent small molecule inhibitors of Leishmania parasites and clinical trials of new drugs are the priorities (**DNDI-0690**, Chart 1).<sup>4,7</sup>

Chart 1. Examples of current antileishmanial medicines and new potent drug candidate.



Beside their antimalarial activity, derivatives of 4aminoquinoline also demonstrated antibacterial, antifungal, antitumor and antileishmanial activity.<sup>8</sup> 4-Amino-7chloroquinoline analogues and their Pt(II) complexes were shown to be active against promastigotes of different *Leishmania* species.<sup>8</sup> Another study investigated the *in vitro* activity of a series of 4-amino-7-chloroquinolines conjugated to sulfonamide, hydrazide and hydrazine against *L. amazonensis* promastigotes and amastigotes, revealing the ability of these Here we report on the synthesis and antileishmanial potential of novel 4-aminoquinoline derivatives and some aminoquinoline-based drugs previously published by our group (Chart 2).<sup>12-16</sup>

#### Chart 2. Structures of examined compounds

1

2

3

4

5

6

7

8

9

10 11

12

13

14

15

16

17

18

19

20

21 22

23

24

25

26

27

28

29

30

31

32

33

34

35

36 37

38

39

40 41

42 43

44 45 46

47

48

49 50

51

52

53

54

55

56

57

58 59

60



The introduction of different substituents at C(3) of quinoline moiety was of interest since it is expected to influence the electronic density to substantial extent. Sontochin-like compounds, as well as our previously published aminoquinoline derivatives with fluorine atom at the same position proved to be of significant antiplasmodial potential.<sup>12</sup> Here, we explore the effect of nitro and amino substituents on antileishmanial activity (Scheme 1). Nitro substituent in **31**<sup>12</sup> enabled nucleophilic substitution under mild conditions,<sup>17</sup> resulting in compounds **1** and **2** in reasonably good yield. Reduction of nitro group to amino using tin(II)-chloride<sup>12</sup> afforded compounds **3** and **4**.

Scheme 1. Synthesis of substituted chloroquine-like compounds 1-4



Aminoquinolines with adamantane carrier were synthesized as presented in Scheme 2. Commercially available 1adamantanemethanol was transformed into mono-Boc protected amine **32** in moderate yield using procedure we established earlier.<sup>12</sup> Removal of protecting group under standard conditions gave amine **33** in good yield. Dichloride **31** was submitted to nucleophilic substitution with amine **33** affording aminoquinoline **5** in good yield. Nitro group in **5** was further reduced to amine **6** using SnCl<sub>2</sub>. Compound **10** with piperazine moiety in linker was synthesized in several reaction steps, starting from commercially available 3,3'-piperazine-1,4-diyldipropan-1-amine **34** (Scheme 3). After protection of amino groups and nucleophilic substitution with 4,7-dichloroquinoline, compound **36** was obtained in moderate yield. Removal of protecting group under standard conditions gave amine **37** in high yield. Reductive amination of the obtained amine with adamantane-1acetaldehyde furnished the compound **38**. Compound **10** was obtained as HCl salt of amine **38** (confirmed by elemental analysis).

Scheme 2. Synthesis of adamantane derivatives 5 and 6



i) 1) PCC, DCM, 2) *tert*-butyl (3-aminopropyl)carbamate, MeOH/DCM, AcOHglac, 3) NaBH<sub>4</sub>;
 ii) TFA, DCM; iii) **31**, DCM, 0 °C to reflux; iv) SnCl<sub>2</sub>, EtOH

Scheme 3. Synthesis of compound 10



Benzothiophene derivative **15** was obtained by reductive amination starting from aldehyde  $39^{13}$  in good yield. Compound **21** was obtained in rather low yield by Sonogashira coupling reaction of  $40^{13}$  and *N*-(prop-2-yn-1-yl)-*N*'-(quinolin-4-yl)butane-1,4-diamine **41**. (Scheme 4).

Scheme 4. Synthesis of novel benzothiophene derivatives 15 and 21



 i) 1) AQ3, AcOHglac, MeOH/DCM, 2) NaBH<sub>4</sub>; ii) 41, PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub>, PPh<sub>3</sub>, Cul, Et<sub>2</sub>NH, DMF, MW, 120 °C, 25 min

Reduction of quinoline core was designed in order to explore the effect of resulting deviation from planarity on inhibition of *Leishmania* proliferation (Scheme 5). Compound **42**, synthesized according to the procedure described in literature,<sup>18</sup> was transformed into **43** in acceptable yield. Selective reduction of benzene core of quinoline and hydrogenolysis of chlorine were performed by hydrogenation method using platinum(IV) oxide in acetic acid as solvent in the presence of

perchloric acid. Deacetylation of compound 43 in 2M HCl gave compound 44 in high yield. Using Sandmeyer reaction conditions (NaNO<sub>2</sub>, AcOH, HCl; CuCl), compound 44 was transformed into 45.

Scheme 5. Synthesis of tetrahydroquinoline core



Syntheses of thiophene-based tetrahydroquinoline compounds are presented on Scheme 6. Compound **45** was submitted to Buchwald-Hartwig amination affording the amines **22**, **23** and **46** in low to moderate yield. Compound **46** with eight methylene groups in linker was subjected to reductive amination with 4-[5-(4-formylphenyl)thiophen-2-yl]benzonitrile<sup>13</sup> to obtain **24**. Methylation of secondary nitrogen using 37% formaldehyde gave compound **25** in high yield.

Scheme 6. Synthesis of compounds 22-25



The syntheses of other compounds are presented in our previous papers.<sup>12-16</sup> Full details of synthetic procedures, NMR spectra and HPLC purities are given in the Supporting Information.

Thirty compounds presented on Chart 2 were first examined for their activity against *L. infantum* and *L. tropica* promastigote stage using standard MTT assay (Table 1, Table S1). Ten compounds showed IC<sub>50</sub> values of the same order of magnitude as amphotericin B (IC<sub>50</sub> < 1  $\mu$ M), which was used as a positive control. C(3)-substituted chloroquine-like compounds (1-4) displayed poor antileishmanial activity against both promastigote species. However, hybrids of such compounds with adamantane carrier resulted in more active derivatives **5** and **6**. Among adamantane derivatives without substituent at C(3), compound **7** showed clear improvement of potency. The most potent compound was **10**, with piperazine moiety in linker.

Benzothiophene compounds 15 and 17 with chlorine atom at C(7) position of quinoline moiety were more potent than their des-chloro analogues 16 and 18, suggesting that chlorine atom would be important for the activity. Replacing phenyl group with C=C bond did not produce any significant effect on the activity (18 vs 21).

While chloroquine-like compounds, tetrahydroquinolines 22 and 23 were completely inactive, hybrids with thiophene carrier 24 and 25 showed >100-fold increase in activity. Among other thiophenes, compound 29 with eight methylene groups between two nitrogens, demonstrated the highest potency against both *Leishmania* promastigote species. All compounds were also checked for cytotoxicity against differentiated THP-1 cells. For both species moderate selectivity indices were obtained ( $SI_{THP-1/L,i}$ =1.3-9.9;  $SI_{THP-1/L,i}$ =1.1-10.9, Table 1).

**Table 1.** *In vitro* activities against *L. infantum* and *L. tropica* promastigotes and cytotoxicity against THP-1 human cells<sup>*a*</sup>

Comp.	L. <i>infantum</i> IC <sub>50</sub> (µM) <sup>b</sup>	L. tropica IC <sub>50</sub> (µM) <sup>b</sup>	ТНР-1 IC <sub>50</sub> (µМ) <sup>с</sup>	SI (THP/ L.i.) <sup>d</sup>	SI (THP/ <i>L.t.</i> ) <sup>d</sup>		
1	8.67	2.77	23.66	2.7	8.5		
2	6.49	2.96	>109.6	>16.9	>37.0		
3	16.60	9.35	>65.2	>3.9	>7.0		
4	16.60	6.63	>59.7	>3.6	>9.0		
5	1.91	2.24	12.59	6.6	5.6		
6	1.77	1.30	6.39	3.6	4.9		
7	0.73	0.66	1.81	2.5	2.7		
8	2.46	1.84	3.29	1.3	1.8		
9	2.40	2.35	4.90	2.0	2.1		
10	0.52	0.51	1.00	1.9	2.0		
11	1.14	1.31	2.96	2.6	2.3		
12	0.64	0.68	3.76	5.8	5.5		
13	1.23	1.24	4.25	3.4	3.4		
14	0.51	0.50	1.91	3.8	3.8		
15	0.48	0.43	4.73	9.9	10.9		
16	1.03	0.81	2.31	2.2	2.8		
17	1.02	0.85	4.28	4.2	5.0		
18	1.24	1.02	2.35	1.9	2.3		
19	0.98	0.91	2.44	2.5	2.7		
20	1.55	1.22	2.79	1.8	2.3		
21	1.02	1.37	7.11	7.0	5.2		
22	>76.5	>76.5	>76.5	>1	>1		
23	>69.1	>69.1	>69.1	>1	>1		
24	0.72	0.75	2.31	3.2	3.1		
25	0.83	0.80	3.68	4.4	4.6		
26	2.30	1.94	5.01	2.2	2.6		
27	1.22	1.54	2.80	2.3	1.8		
28	5.42	7.11	8.10	1.5	1.1		
29	0.35	0.30	1.38	4.0	4.6		
30	0.80	1.06	3.85	4.8	3.6		
Control <sup>e</sup>	0.13	0.14	>10.8	>83.1	>77.1		
aAntileishmanial IC50 values against promastigote stages (µM), MTT assay; bAll <i>in vitro</i> experiments were performed in duplicate, mean values are given; cCytotoxicity against differentiated THP-1, hu- man monocytic cell line derived from an acute monocytic leukemia patient. dSelectivity index; eControl drug: amphotericin B							

All compounds were tested against intramacrophage amastigotes of *L. infantum* at 0.5  $\mu$ M, non-toxic concentration on human cells (Table S2). Compounds that showed more than 25% inhibition were tested in dose-response experiments and IC<sub>50</sub> were calculated. Five compounds showed IC<sub>50</sub> less than 1  $\mu$ M. Among them, compounds **10**, **15** and **18** were the most active, while compound **15** was the least toxic with good selectivity index (Table 2).

Three compounds with good antileishmanial potential (10, 15 and 18) were subjected to *in vivo* tolerability evaluation in a mice model. All compounds were tested orally at 300 mg/kg (single dose) as suspension in 0.1%Tween/0.5% HEC in wa-

1

2

3 4

5

6 7

8 9

10

11

12 13

14

15

16

17

18

19

20 21

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57 58 59

60

ter. Compound **15** was also tested at lower 50 mg/kg dose by subcutaneous route of administration (in sunflower oil). Compound **18** showed toxic effects when given orally with 3/5 mice alive at the end of experiment, while compounds **10** and **15** given either p.o. or s.c. proved to be tolerable, since all mice survived 30 days after administration and showed normal appearance and behavior.

 
 Table 2. In vitro activities against intramacrophage L. infantum amastigotes

Comp.	In Vitro Antiamastigote Activity at 0.5 µM <sup>a</sup>	In Vitro Antiamastigote Activity IC <sub>50</sub> (µM) <sup>b</sup>	THP-1 <sup>c</sup> IC <sub>50</sub> (μM)	SI (THP/IPT) <sup>d</sup>
8	29.6	1.91	3.29	1.72
10	72.2	0.31	1.00	3.22
11	26.4	1.85	2.96	1.60
13	38.9	1.29	4.25	3.29
14	26.4	>1	1.91	<1.91
15	47.6	0.58	4.73	8.15
18	42.7	0.65	2.35	3.61
19	36.9	0.73	2.44	3.34
20	42.2	0.79	2.79	3.51
24	29.6	>1	2.31	<2.31
Control <sup>e</sup>	95.5	0.21	>10.8	>51.4

aMean value of two or three experiments. bMean value of two experiments. cCytotoxicity against differentiated THP-1, human monocytic cell line derived from an acute monocytic leukemia patient. dSelectivity Index (IC50 against THP-1/IC50 against intracellular amastigotes); eControl drug; amphotericin B.

Compounds 10 and 15 were further evaluated for reduction of liver parasite load in a mouse model of visceral leishmaniasis (Balb/c mice infected intravenously with L. infantum amastigotes). Results presented in Figure 1 are given as % of reduction relative to control (untreated infected mice, Table S3, Table S4). Compound 15 was tested per os at two different doses 50 mg/kg  $\times$  4 days and 100 mg/kg  $\times$  4 days and showed significant reduction of parasites in the liver, 95% and 99%, respectively. Compound 10 was also tested per os at 60 mg/kg  $\times$  4 days and 100 mg/kg  $\times$  4 days. At lower applied dose it reduced parasite load 96% compared to control. Although at higher dose complete clearance was achieved, it showed signs of toxicity, since 1 mouse died on day 10 and 1 mouse died on day 12. Both compounds were also subjected to s.c. administration at lower doses (Figure 1). Compound 10 administered at dose 10 mg/kg  $\times$  4 days reduced parasite load by 81%. At  $5 \text{ mg/kg} \times 5 \text{ days and lower both compounds exhibited poorer}$ activities (57% and 18-48% for 10 and 15, respectively). However, these results are extremely important, giving us the essential information about dose-dependent behavior of selected 4-aminoquinolines.

In order to investigate the putative mechanism of action (MOA),<sup>9</sup> we examined the production of nitric oxide and ROS by IFN $\gamma$  primed murine bone marrow-derived macrophages (BMDM) treated with **10** or **15**. Experiments were performed at several concentrations not toxic on BMDM. Not toxic concentrations were determined by MTT assay (data not shown). The concentration of nitrite and ROS was determined using Griess reagent and H<sub>2</sub>DCFDA, respectively. Results revealed that compounds **10** and **15** increased the production of nitric oxide by IFN $\gamma$ -primed macrophages only at the highest dose used (Figure S1). Both compounds induced persistent increase of ROS at all the doses tested (Figure S2).



Page 4 of 6

Figure 1. Reduction of parasite load in a mouse model by compounds 10 and 15

Currently, several noteworthy *in vivo* studies have been published,<sup>19,20,21</sup> Nitroimidazo-oxazole compound **DNDI-VL-2098** previously identified as a favorable candidate did not proceed to the clinical study.<sup>20,22</sup> However, its oxazine derivative **DNDI-0690** was very recently recognized as a new promising candidate for Phase I trial for VL.<sup>4,7</sup> In the last ten years, only a few 4-amino- and 2-alkenylquinolines with modest activity against *Leishmania* parasites *in vivo* were also identified.<sup>23-25</sup> 8-Aminoquinoline derivative sitamaquine appeared to be orally active against visceral leishmaniasis and is presently under clinical investigation.<sup>26</sup>

Here, we identified two 4-amino-7-chloroquinoline compounds bearing benzothiophene or adamantane moieties as potent antileishmanial candidates with significant *in vivo* efficacy. The importance of this work lies in discovery of highly active short-term compounds, tolerable in mice, being the first example of a 4-amino-7-chloroquinoline active in *L. infantum* mice model of visceral leishmaniasis. Although a certain dosedependent enhancement of NO and ROS production (which can contribute to the amastigotes killing) was observed in the presence of **10** and **15**, MOA still remains unclear. Further studies will be focused on discovering specific target in order to elucidate the mechanism of action. Based on compounds' tolerability in mice model and noteworthy *in vivo* activity, future efforts will be focused on improvement of pharmacokinetic profile and enhancement of the antileishmanial activity.

#### ASSOCIATED CONTENT

#### **Supporting Information**

The Supporting Information is available free of charge on the ACS Publications website.

Full details of synthetic procedures, biological assays, procedures for the determination of the HPLC purity, *in vitro* activities on promastigote stages (Table S1), *in vitro* activities against intramacrophages *L. infantum* amastigotes (Table S2), *in vivo* antileishmanial activity (Table S3, Table S4), nitric oxide and ROS production (Figure S1, Figure S2) (**Supporting information – I**, PDF).

NMR spectra and HPLC purity spectra of all tested compounds (**Supporting information – II**, PDF).

#### AUTHOR INFORMATION

**Corresponding Author** 

59

60

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58 59

60

\*For B.Š.: phone, +381-11-263-86-06; fax, +381-11-263-60-61;

E-mail: <u>bsolaja@chem.bg.ac.rs; bogdan.solaja@sanu.ac.rs</u>.

\*For N.B.: phone, +39-02 5031-5069; fax: +39-02-5031-5093;

E-mail: nicoletta.basilico@unimi.it.

#### Author Contributions

B.Š. and N.B. designed the research. The manuscript was written by J.K. with contributions of all authors. All authors have given approval to the final version of the manuscript.

#### Funding Sources

This work was supported by the Ministry of Science and Technological Development of Serbia (Grant 172008), Serbian Academy of Sciences and Arts, Executive Programme of Scientific and Technological Cooperation between the Italian Republic and the Republic of Serbia for the years 2016-2018 and by "Ministero dell'Istruzione, dell'Università e della Ricerca (PRIN) Project: 20154JRJPP\_004".

#### Notes

The authors declare no competing financial interest.

*Ethical approval:* The study followed the International Guiding Principles for biomedical research involving animals (European Directive 2010/63/UE), and it was reviewed by a local Ethics Committees. The study was approved by the Veterinary Directorate at the Ministry of Agriculture and Environmental Protection of Serbia (decision no. 323-07-02444/2014-05/1) and by the Directorate of Animal Health and Veterinary Drugs at the Ministry of Health of Italy (authorization no. 120/2015-PR).

#### ACKNOWLEDGMENT

We thank Dr. Olgica Djurković-Djaković and MSc Jelena Srbljanović (Institute for Medical Research, University of Belgrade) for performing *in vivo* toxicity studies and Loredana Cavicchini for assistance in culturing leishmania *in vitro*. We also thank Prof. Donatella Taramelli (Dipartimento di Scienze Farmacologiche e Biomolecolari, University of Milan) for helpful discussion, and COST Action CM1307 for support.

#### ABBREVIATIONS

BMDM, murine bone marrow-derived macrophages; MTT, 3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide; Tween 80, Polysorbate 80; HEC, hydroxyethyl cellulose; ROS, reactive oxygen species; MOA, mechanism of action; **AQ3**, *N*-(7chloroquinolin-4-yl)propane-1,3-diamine.

#### REFERENCES

1. Leishmaniasis, Fact Sheet N°375, February 2015. http://www.who.int/mediacentre/factsheets/fs375/en/ (accessed October 16, 2017).

2. Centers for Disease Control and Prevention, Parasites - Leishmaniasis, January 2013. http://www.cdc.gov/parasites/leishmaniasis/ (accessed October 16, 2017).

3. WHO Technical Report Series no. 949, Report of a Meeting of the WHO Expert Committee on the Control of Leishmaniases, Geneva, 22–26 March 2010. http://www.who.int/neglected\_diseases/resources/who\_trs\_949/en/

(accessed October 16, 2017).

4. https://www.dndi.org/diseases-projects/portfolio/ (accessed March 22, 2018).

5. Sangshetti, J. N.; Kalam Khan, F. A.; Kulkarni, A. A.; Aroteb, R.; Patil, R. H. Antileishmanial drug discovery: comprehensive review of the last 10 years. *RSC Adv.* **2015**, *5*, 32376-32415.

6. Gillespie, P. M.; Beaumier, C. M.; Strych, U.; Hayward, T.; Hotez, P. J.; Bottazzi, M. E. Status of Vaccine Research and Development of Vaccines for Leishmaniasis. *Vaccine* **2016**, *34*, 2992-2995.

7. Thompson, A. M.; O'Connor, P. D.; Marshall, A. J.; Yardley, V.; Maes, L.; Gupta, S.; Launay, D.; Braillard, S.; Chatelain, E.; Franzblau, S. G.; Wan, B.; Wang, Y.; Ma, Z.; Cooper, C. B.; Denny, W. A. 7-Substituted 2-Nitro-5,6-dihydroimidazo[2,1-b][1,3]oxazines: Novel Antitubercular Agents Lead to a New Preclinical Candidate for Visceral Leishmaniasis. *J. Med. Chem.* **2017**, *60*, 4212-4233.

8. Carmo, A. M. L.; Silva, F. M. C.; Machado, P. A.; Fontes, A. P. S.; Pavan, F. R.; Leite, C. Q. F.; Leite, S. R. de A.; Coimbra, E. S.; Da Silva, A. D. Synthesis of 4-Aminoquinoline Analogues and Their Platinum(II) Complexes as New Antileishmanial and Antitubercular agents. *Biomed. Pharmacother.* **2011**, *65*, 204–209. and ref. therein.

9. Antinarelli, L. M. R.; Dias, R. M. P.; Souza, I. O.; Lima, W. P.; Gameiro, J.; da Silva, A. D.; Coimbra, E. S. 4-Aminoquinoline Derivatives as Potential Antileishmanial Agents. *Chem. Biol. Drug Des.* **2015**, *86*, 704-714.

10. Soares, R. R.; Antinarelli, L. M. R.; Souza, I. O.; Lopes, F. V.; Scopel, K. K. G.; Coimbra, E. S.; da Silva, A. D.; Abramo, C. In Vivo Antimalarial and In Vitro Antileishmanial Activity of 4- Aminoquinoline Derivatives Hybridized to Isoniazid or Sulfa or Hydrazine Groups. *Lett. Drug Des. Discov.* **2017**, *14*, 597 – 604.

11. Antinarelli, L. M. R.; Carmo, A. M. L.; Pavan, F. R.; Leite, C. Q. F.; Da Silva, A. D.; Coimbra, E. S.; Salunke, D. B. Increase of Leishmanicidal and Tubercular Activities Using Steroids Linked to Aminoquinoline. *Org. Med. Chem. Lett.* **2012**, *2*, 16.

12. Terzić, N.; Konstantinović, J.; Tot, M.; Burojević, J.; Djurković-Djaković, O.; Srbljanović, J.; Štajner, T.; Verbić, T.; Zlatović, M.; Machado, M.; Albuquerque, I. S.; Prudêncio, M.; Sciotti, R. J.; Pecic, S.; D'Alessandro, S.; Taramelli, D.; Šolaja, B. A. Reinvestigating Old Pharmacophores: Are 4-Aminoquinolines and Tetraoxanes Potential Two-Stage Antimalarials? *J. Med. Chem.* **2016**, *59*, 264 – 281.

13. Konstantinović, J.; Videnović, M.; Srbljanović, J.; Djurković-Djaković, O.; Bogojević, K.; Sciotti, R.; Šolaja, B. Antimalarials With Benzothiophene Moieties as Aminoquinoline Partners. *Molecules* **2017**, *22*, 343.

14. Konstantinović, J.; Kiris, E.; Kota, K.; Kugelman-Tonos, J.; Videnović, M.; Cazares, L. H.; Terzić, N.; Verbić, T. Ž.; Andjelković, B.; Duplantier, A. J.; Bavari, S.; Šolaja, B. A. New Steroidal 4-Aminoquinolines Antagonize Botulinum Neurotoxin Serotype A in Mouse Embryonic Stem Cell Derived Motor Neurons in Post-intoxication Model. *J. Med. Chem.* **2018**, *61*, 1595-1608.

15. Marković, O. S.; Cvijetić, I. N.; Zlatović, M. V.; Opsenica, I. M.; Konstantinović, J. M.; Terzić Jovanović, N. V.; Šolaja, B. A.; Verbić, T. Ž. Human Serum Albumin Binding of Certain Antimalarials. *Spectrochim. Acta Mol. Biomol. Spectrosc.* **2018**, *192*, 128-139.

16. Šolaja, B. A.; Opsenica, D.; Smith, K. S.; Milhous, W. K.; Terzic, N.; Opsenica, I.; Burnett, J. C.; Nuss, J.; Gussio, R.; Bavari, S. Novel 4-Aminoquinolines Active against Chloroquine-Resistant and Sensitive P. falciparum Strains that also Inhibit Botulinum Serotype A. J. Med. Chem. **2008**, *51*, 4388–4391.

17. Gerster, J. F.; Lindstrom, K. J.; Miller, R. L.; Tomai, M. A.; Birmachu, W.; Bomersine, S. N.; Gibson, S. J.; Imbertson, L. M.; Jacobson, J. R.; Knafla, R. T.; Maye, P. V.; Nikolaides, N.; Oneyemi, F. Y.; Parkhurst, G. J.; Pecore, S. E.; Reiter, M. J.; Scribner, L. S.; Testerman, T. L.; Thompson, N. J.; Wagner, T. L.; Weeks, C. E.; Andre, J.-D.; Lagain, D.; Bastard, Y.; Lupu, M. Synthesis and Structure–Activity-Relationships of 1H-Imidazo[4,5-c]quinolines That Induce Interferon Production. *J. Med. Chem.* **2005**, *48*, 3481–3491.

18. Korotchenko,V.; Sathunuru,R.; Gerena, L.; Caridha, D.; Li, Q.; Kreishman-Deitrick, M.; Smith, P. L.; Lin, A. J. Antimalarial Activity of 4-Amidinoquinoline and 10- Amidinobenzonaphthyridine Derivatives. *J. Med. Chem.* **2015**, *58*, 3411-3431.

19. Dea-Ayuela, M. A.; Castillo, E.; Gonzalez-Alvarez, M.; Vega, C.; Rolón, M.; Bolás-Fernández, F.; Borrás, J.; González-Rosende, M. E. In Vivo and in Vitro Anti-leishmanial Activities of 4-Nitro-N-

pyrimidin and N-Pyrazin-2-ylbenzenesulfonamides, and N2-(4-Nitrophenyl)-N1-propylglycinamide. Bioorg. Med. Chem. 2009, 17, 7449-7456.

20. Gupta, S.; Yardley, V.; Vishwakarma1, P.; Shivahare1, R.; Sharma, B.; Launay, D.; Martin, D.; Puri, S. K. Nitroimidazo-oxazole Compound DNDI-VL-2098: An Orally Effective Preclinical Drug Candidate for the Treatment of Visceral Leishmaniasis. J. Antimicrob. Chemother. 2015, 70, 518-527.

21. Galiana-Roselló, C.; Bilbao-Ramos, P.; Dea-Ayuela, M. A.; Rolón, M.; Vega, C.; Bolás-Fernández, F.; García-España, E.; Alfonso, J.; Coronel, C.; González-Rosende, M. E. In Vitro and In Vivo Antileishmanial and Trypanocidal Studies of New N-Benzene- and N-Naphthalenesulfonamide Derivatives. J. Med. Chem. 2013, 56, 8984-8998.

22. https://www.dndi.org/diseases-projects/portfolio/completedprojects/vl-2098/ (accessed March 22, 2018).

23. Nakayama, H.; Desrivot, J.; Bories, C.; Franck, X.; Figade're, B.; Hocquemiller, R.; Fournet, A.; Loiseau, P.M. In Vitro and In Vivo Antileishmanial Efficacy of a New Nitrilquinoline against Leishmania donovani. Biomed Pharmacother. 2007, 61, 186-188.

24. Gopinath, V.S.; Pinjari, J.; Dere, R. T.; Verma, A.; Vishwakarma, P.; Shivahare, R.; Moger, M.; Kumar Goud, P. S.; Ramanathan, V.; Bose, P.; Rao, M. V.; Gupta, S.; Puri, S. K.; Launay, D.; Martin, D. Design, Synthesis and Biological Evaluation of 2-Substituted Quinolines as Potential Antileishmanial Agents. E. J. Med. Chem. 2013, 69, 527-536.

25. Deshpande, S.; Shivahare, R.; Debnath, U.; Shraddha, A. S.; Gupta, S.; Seturam, B. K. Synthesis and Bio-evaluation of 7trifluromethyl Substituted 4-aminoquinoline Derivatives as Antileishmanial Agents. The Natural Products Journal 2017, 7, 137-143.

26. Loiseau, P. M.; Cojean, S.; Schrével, J. Sitamaquine as a Putative Antileishmanial Drug Candidate: From the Mechanism of Action to the Risk of Drug Resistance. Parasite 2011, 18, 115-119.

